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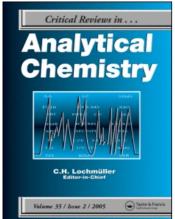
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Trace Analysis — Challenges and Problems

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ABSTRACT: Analytical procedures and techniques employed to determine trace components are objects of growing interest to analysts. The determination of those components in samples of different matrix composition is becoming ever more common. Hence, attention should be paid to problems involved in this type of activity.

The article presents specific problems connected with trace analysis. The problems and solutions proposed are documented with data from the literature.

KEY WORDS: trace analysis, sources of error, prevention, wall memory effect, cross-contamination.

I. INTRODUCTION

Even a cursory perusal of any analytical journal must lead one to the conclusion that trace and ultra-trace analysis is a domain of chemical analysis that is gaining in importance recently. This conclusion is corroborated not only by the feelings and opinions of analysts. By the current definition of the term 'trace component' proposed by the IUPAC, the limit from which we can talk about trace analysis is the concentration of 100 ppm (100 μ g/g). Naturally, this limit is purely conventional and is not a constant. As recently as 30 years ago 'trace analysis' was understood to denote activities aiming to determine components at a concentration level one order of magnitude higher (i.e., below 1000 ppm, or 0.1%).

Even today the determination of components at a concentration level of 100 ppm, even in samples with complex matrices, does not pose major problems and is done routinely in many laboratories. This is mainly due to the rapid development of instrumen-

tation, or the science of the construction and use of monitoring and measuring devices. Hence, one may expect the definition of the term 'trace component' to change again soon. Table 1 presents a classification of analytical methods and techniques by analyte concentration in the sample examined.

II. AREA OF APPLICATION OF TRACE ANALYSIS

It is possible to distinguish three areas of science and technology that spur the development of analytical methods and techniques employed in the determination of low and very low analyte contents in samples of various kinds. They are

• technologies of the production of highpurity materials; to date, the purity of the cleanest man-made material is denoted by 11 N, which means that the sum total of all impurities it contains does not exceed 10⁻⁹%, or 10 ppt;

Table 1. Classification of analytical methods and techniques by analyte concentration in a sample.

General name of analyte	Analyte concentration	Common term for analytical procedure	Examples
Sub-microtrace component	< 1 ppt (< 10 ⁻⁸ %)		Determination of dioxins in samples of various matrices
Ultra-microtrace component	< 1 ppb (< 10 ⁻⁶ %)	TRACE ANALYSIS	Determination of trihalomethanes in drinking water and human urine. Determination of volatile organic compounds in indoor air
Microtrace component	< 1 ppm (< 10 ⁻⁴ %)		Determination of carbon monoxide in ambient air
Trace component	< 100 ppm (< 0.01%)		Determination of methane in ambient air
Secondary component (admixture)	< 1%	SEMI- MICROANALYSIS	Determination of carbon dioxide in ambient air
Primary component	1-100%	MACROANALYSIS	Determination of oxygen in waste gases. Determination of oxygen in flue gases

- genetic engineering and biotechnology;
- environmental protection.

The determination of ever lower concentrations of analytes has brought into common use special ways of expressing such concentrations. Table 2 lists the units employed to denote concentrations in trace analysis.

Efforts to determine analytes at ever lower concentration levels in samples with complex matrices that additionally exhibit a high degree of nonhomogeneity are the most important tendency in the development of modern chemical analysis.^{1,2} Advances in this field are connected primarily with three types of activity:

- the use in analytical practice of new kinds of detectors and sensors (including biosensors) characterised by:
 - a low threshold of detectability and determinability

- —high selectivity, and often even specificity of response;
- the designing of techniques of sample preparation for the stage of final determination.
 There is no doubt that this avenue of research opens up especially many new possibilities of determination of trace and ultratrace components. Table 3 lists the main tasks of the stage of sample preparation for analysis;
- the use of hyphenated techniques in analysis. Their development is connected with the growing significance of speciation analysis. Complex analytical systems have been devised which are often composed of three elements connected on-line:³⁻⁶
 - a system for preliminary sample preparation, usually involving the technique of extraction to the solid phase (Figure 1 shows a diagram of the basic stages of sample preparation using the SPE technique);
 - —a separation system. Here a special role is played by chromatographic methods

Table 2. Units employed to denote concentrations of trace and ultra-trace components.

Name of concentration unit	Part per thousand	Part per million	Part per billion	Part per trillion	Part per quadrillion	Part per quintillion	Part per sextillion
Volume/volume concentration	vpth (ppth v/v)	(v/v mdd)	(v/v dqq)	vpt (ppt v/v)	(n/n bdd) bdn	vpq _{ui} (v/v)	(v/v sqq)
Mass-mass concentration	ppth	шдд	qdd	ppt	bdd	ppd ⁿⁱ	sdd
Percentage (%)	10-1	10-4	10-7	10-10	10-13	10-16	10-19
Amount of analyte in 1 g sample	1 milligram (1 mg)	1 microgram (1 µg)	1 nanogram (1 ng)	1 picogram (1 pg)	1 femtogram (1 fg)	1 attogram (1 ag)	1 zeptogram (1 zg)

- (mainly GC and HPLC). They are usually employed to separate mixtures into individual components;
- —a suitable detection system. For example, Figure 2 presents combinations of separation techniques and spectroscopic detection systems that are most widely used in analytical practice.⁶

III. MAIN SOURCES OF ERROR IN TRACE ANALYSIS

Obviously enough, the determination of components that occur in a sample at very low concentrations presents a completely different challenge to analysts. The various operations performed on the sample during its preparation for the stage of final determinations can be a source of many errors crucially affecting the final result of the analysis.

The chief sources of error are changes in the composition of the sample under study brought about by a variety of factors. In Table 4 three principal kinds of error distorting the result of analysis are presented.

The bias of the final determination result caused by the errors listed (Table 4) may be due to various processes occurring in the sample and due to the influence of the external environment on the sample. By way of example, Figure 3 presents diagrammatically the sources of error that can distort the result of determination of a trace component in a liquid sample. The intensity of the processes and phenomena shown in the figure can vary depending on:

- the temperature of the sample and its changes,
- the presence and intensity of solar radiation, and
- the presence of living organisms in the sample.

The contact of analytes present in both gas and liquid mixtures with the walls of vessels, tubing and appliances crucially affects the concentration levels of trace and ultratrace components. It is the result of adsorption and desorption of analytes on the surface of a solid. These phenomena distorting the sample composition are described collectively as 'the wall memory effect' and 'cross-contamination'. Table 5 shows diagrammatically the magnitude of polar organic analyte-wall surface interaction.

What gives rise to many misunderstandings is the use in analysis of water of suitable purity. Table 6 lists data concerning the efficiency of some water-purifying methods, while Table 7 gives specific electrical conductance values for water with various degrees of purity.

Obtaining ultrapure water, which is the basic reagent in laboratories performing trace analysis, is usually a multistage process. In the installation for the production of water of this level of purity the following processes are employed:

- preliminary filtering to remove most suspended matter;
- activated-carbon filtering to remove free chlorine (to a sub-1-ppm level), small particles of suspended matter (1 to 5 μm) and most organic matter (to a level below 1 ppm of carbon);
- preliminary chemical processing, consisting in the neutralization of water with acid or alkalies, as required, and in the transformation of dissolved CO₂ into carbonates and bicarbonates;
- ionic exchange, during which ions present in water, including those introduced in the processing, are exchanged for hydrogen and hydroxyl ions on the cation and anion exchangers;
- membrane filtration microfiltration, ultrafiltration, reverse osmosis;

Table 3. Stages of sample preparation for final determinations.

Task	Way of implementation
Ensuring sample stability during transport and storage	Chemical conservation Thermal conservation Lyophilization
Sample homogenization	Grounding Mixing Screening
Removal of interfering components	Dust removal (gaseous samples) Suspension removal (liquid samples) Sample drying Sample deoxidation Removal of reactive components
Chemical conversion of analytes (derivatization) into forms easy to: - isolate - separate mixture components - detect and quantitate	In situ derivatization (while sampling analytes) Derivatization in column Derivatization in eluate
Exchange of sample matrix for one 'friendly' to measuring device used	Extraction of analytes from sample using: - stream of washing gas - suitable solvent (including supercritical fluids) - membrane devices - thermal desorber
Increasing analyte concentration in examined sample to level enabling quantitative analysis	Use of wide variety of analyte enrichment techniques (in many cases this is connected with matrix exchange)
Reducing amount of reagents used (including solvents)	Use of so-called solvent-free techniques of sample preparation Reduction of determination scale Introduction of Total Chemical Analysis System (<i>TAS</i>) into analytical practice

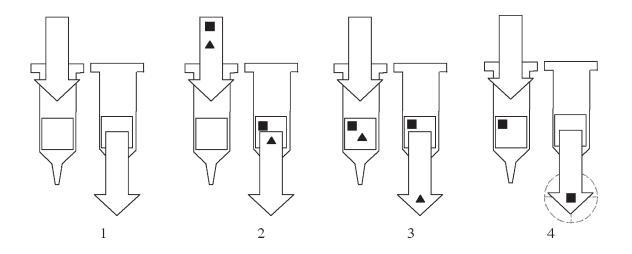


FIGURE 1. Diagram of the main stages of extraction to the solid phase.

1 - conditioning of sorbent bed, 2 - adsorption (passing stream of sample through sorbent bed), 3 - rinsing and cleaning of sorbent bed, 4 - elution of analytes (obtaining of solvent concentrate)

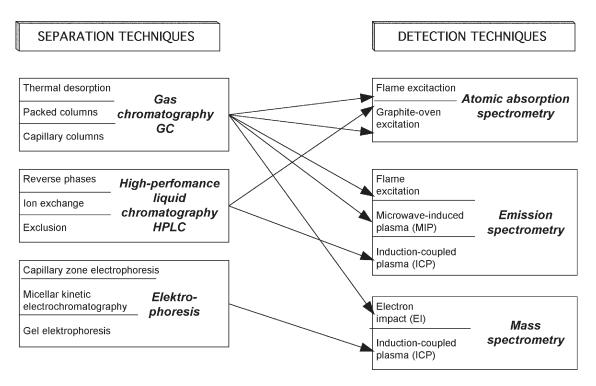


FIGURE 2. Combination of separation techniques and spectroscopic detection techniques most widely used in analysis.

Table 4. Sources of error affecting the ultimate result of analysis.

Source of error	Name of error
"Delivery" of additional quantities of analyte to sample	Positive error
Loss of analyte	Negative error
"Appearance" of additional components in sample	Interferences

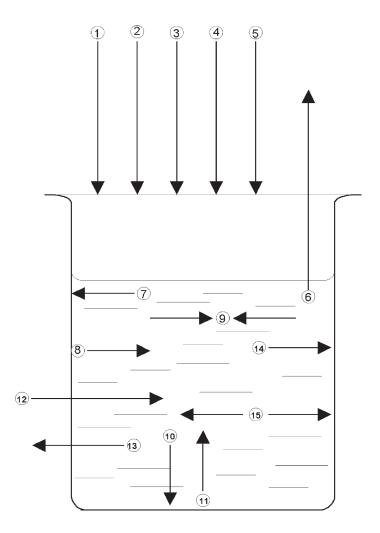


FIGURE 3. Diagrammatic presentation of factors that can affect concentration levels of trace components in a liquid sample.

1 - contact of sample with laboratory air; 2 - residues of components of dishwashing mixtures; 3 - distilled water; 4 - reagents and solvents employed; 5 - contact with analyst; 6 - vaporization of the most volatile components; 7-8 - processes of adsorption-desorption (wall memory effect); 9 - adsorption of analytes on suspension; 10 - precipitation of sediment; 11 - leaching of components from vessel; 12-13 - permeation of solution components through vessel walls; 14 - reaction of analyte with vessel material; 15 - chemical reactions among solution components.

Table 5. Comparison of the polar organic analyte-wall surface interaction for various construction materials and ways of preparing vessel walls.

Construction material	Magnitude of analyte-wall interaction
Chromium-molybdenum steel Teflon-coated steel Aluminium Glass (after washing with acid) Aluminium (after surface cleansing) Electropolished stainless steel Electropolished stainless steel (after surface cleansing) Glass (after silanization)	g)

Table 6. Comparison of the efficiency of some water-purifying methods.

		Pollute	ant	
Water-purifying techniques	colloids	organic substances	ions	bacteria
Sorption on activated carbon	0	1	0	0
Reverse osmosis	3	2	1	3
Ionic exchange	1	1	3	0
Filtering through filter with pores 0.2 µm in diameter	3	0	0	3

- 0 unsatisfactory purification
- 1 satisfactory purification
- 2 good purification
- 3 very good purification

Table 7. Comparison of specific electrical conductance values of raw, distilled and deionized water.

Kind of water	Specific conductance, μS/cm
Raw water	300 - 1000 and over
Distilled water: - distilled once - distilled twice from quartz - distilled twice from glass	4 - 6.5 0.5 1.0
Deionized water	0.2 - 0.1 or under (to 0.055)

- water degassing to remove oxygen and carbon dioxide, and to reduce the content of volatile organic compounds to a level below 1 ppb;
- UV sterilization, which ensures a reduction of bacteria in water to a level below 1 bacterium/100 cm³ of water, and the oxygenation of the remaining part of organic matter to a level below 1 ppb;
- water ozonation to further reduce the content of organic matter.

For years the amount of organic matter in water (usually expressed by the total content of organic carbon) has been considered the chief measure of quality of high-purity water.

Analytic work in the field of trace analysis should be carried out not only with the use of highest-purity reagents, but also in rooms conforming to the 'clean room' standards. Such a room is a space separated from ambient air and only accessible through a sluice. A stream of air cleaned by a system of filters is pumped into the room under a slight positive gauge pressure. The excess is removed through a lattice floor. The HEPA filter that is part of the system stops dust particles of more than 0.3 µm with an efficiency of up to

99.99%. With the air cleaned in this way, controlled humidity and temperature, and suitable working rules of the personnel, the risk of sample contamination is reduced to a minimum in such a laboratory. There has appeared a first mention of the so-called ultraclean chemical laboratory (UCCL) in the literature. In its case the air quality must meet even more stringent criteria. Unfortunately, a significant limitation to 'clean rooms' is the very high cost of such an investment. That is why it is much more common for laboratories to utilize much cheaper 'clean boxes' with a laminar flow of an inert gas. They can be used successfully in sample processing. The introduction of the robotization and automation of sample processing into analytical practice may be a solution to many problems connected with the cleanness of the laboratory environment.

The quality of the air is defined through the number of dust particles in a unit volume. The American standards set in the U.S. Federal Standard 209 distinguish six classes of laboratory air on the basis of the number of particles from 0.5 to 5.0 μ m in diameter contained in 28.32 dm³ of air. The classification is presented in Table 8.

In Table 9 some measures are presented which can eliminate, or at least reduce the

Table 8. Cleanness classes of laboratory air. The number of particles in 28.32 dm³ of air.

Particle diameter Class of air cleanness	0.1	0.2	0.3	0.5	1.0	5.0
1	35	7	3	1	-	-
10	350	75	30	10	-	-
100	-	750	300	100	-	-
1,000	-	-	-	1,000	-	7
10,000		-	-	10,000	-	70
100,000	-	-	-	100,000	-	700

Table 9. Ways of eliminating or reducing the influence of factors affecting concentration levels of trace components in a liquid sample (see Fig. 2).

Factor affecting concentration of trace component in liquid sample	Countermeasures
Contact of sample with laboratory air	- making all activities and operations air-tight - use of clean boxes and clean rooms to carry out operations involved in sample preparation for analysis
Residues of components of dishwashing mixtures	use of proper dishwashing agents and suitable (tested) procedures of cleaning, washing and drying of vessels
Water used in sample preparation operations	proper techniques of water preparation (deionization, distillation, etc.)
4. Reagents and solvents employed	- use of high purity reagents (HPRs) - use of reagents from same manufactured unit - addition of reagents in justified excess - reducing scale of determinations - use of so-called solvent-free techniques of sample preparation
5. Contact with analyst	- use of protective clothes (headgear, gloves, etc.)
6. Vaporization of volatile components	making sample preparation operations air-tight storage of solutions and samples in vessels filled up to stopper use of vessels of appropriate capacity
7-8. Processes of adsorption-desorption of trace components on walls (wall memory effect)	- use of vessels made of suitable materials - special preparation of vessel surface (deactivation) through: - electropolishing - electropassivation - silanization - lowering of storage temperature of samples and solutions - rinsing of vessels with bit of sample or solution
Adsorption of components (primarily organic analytes) on surface of teflon-coated mixer	- another way of mixing sample - use of glass-coated mixer
10. Adsorption of analytes on suspension	- preliminary suspension removal through: - decanting - filtration - centrifugation
11. Precipitation of sediment	- acidification of sample
12. Leaching of components from vessel material	- use of vessels made of suitable materials

Table 9 (continued)

13. Permeation of air components to solution	- use of vessels made of materials of low permeability to gases
14. Permeation of solution components outside	use of vessels made of materials of low permeability to solution components use of thick-walled plastic vessels
15. Reaction of analytes with vessel material	- special preparation of vessel surfaces as in points 7-
Chemical reactions among solution components	- lowering of solution temperature - preliminary sample preparation through derivatization of reactive components
17. Photodegradation	- storage of samples in the dark
18. Biodegradation	- addition of biocides

intensity of, factors likely to affect the concentration levels of trace components in a liquid sample. In the case of gaseous or solid samples, there will appear not only their specific sources of error, but also those (or at least some of those) characteristic of liquid samples.

In the literature one can find plenty of more detailed information about sources of error in trace analysis. There have appeared comprehensive overviews devoted to these issues⁷⁻¹⁰. They deal primarily with inorganic trace analysis.

The available literature also supplies much information (of various levels of detail) concerning problems and sources of error involved in the determination of trace components in samples with complex matrices. In Table 10 the relevant information is listed that concerns trace analysis, while Table 11 provides data about specific issues related to trace analysis in three basic types of sample (gaseous, liquid, and solid).

SUMMARY

Trace analysis is an important part of chemical analysis and is becoming an object of interest to ever growing ranks of analysts. In solving analytical tasks in this field, more and more complicated monitoring and measuring equipment is used. Unfortunately, this does not always go hand in hand with the level of knowledge of the personnel that operates such devices. There have been a growing number of cases when an analytical device is treated like a typical black box into which it suffices to put a sample to get the required analytical information. This approach is, of course, false and the determination results obtained in this way are often a source of misinformation. The author hopes that the present article will be useful in pointing out several problems relevant in determining components occurring at low and very low concentrations in samples with complex compositions.

Table 10. Information about sources of error in trace and ultra-trace analysis.

Source of error	Additional information	Source
	Purity of substances used in trace analysis	=
	Contamination sources on sampling high-purity reagents	12
	Stability of standard aqueous solutions of organic substances - interlaboratory test	13
Reagents used	Purity of acids used in electronic industry	14
	Purity of water, acids and various materials used in trace analysis	15
	Use of ultra-pure water in trace analysis and high-purity materials industry	[16-18]
Condition and equipment of laboratory	Use of clean rooms in trace analysis	[19-21]
	Phenomena affecting representativeness of groundwater samples	22
Determination of sample size	Determination of size of representative sample	23
	Water penetration through walls of vessels made of plastics	24
	Sources of error in trace analysis	25
	Wall memory effect in trace analysis	26
	Effect of various parameters on results of study of degeneration of organic compounds in groundwater	27
1	Occurrence of organic film on surfaces of solids and liquids	28
taking, storage and preparation of	Accuracy in speciation analysis. Use of isotope dilution mass spectrometry (IDMS)	59
samples for analysis	Sources of error involved in taking, storage and analysis of water samples for metallic content	30
	Effect of humic substances on efficiency of reverse osmosis and nanofiltration	31
	Rules of preparation of solid material samples for examination using atomic spectroscopy	32
	Effect of particular steps of analytical procedure on results of determination of trace amounts of elements	33
	Effect of permeation dryer on operation of analytical device (ICP-AES)	34
Measurement consistency	Organisation of international measuring system	35

Source [40-41] 43 36 37 ထ္တ 39 42 44 45 46 47 48 6 22 23 54 20 22 26 28 2 57 Dependence of stability of gaseous samples containing odorous substances on analyte concentrations Changes in adsorption of analytes on bed of XAD-2 sorbent as result of presence of HCl in gas stream Effect of filtration of (dust removal from) gas streams on concentrations of volatile organic compounds Formation of artifacts during sampling of carbonyl compounds - passing of air stream through sorption Effect of construction of sampler and measuring set on assay results of analytes from PCDD/F group Changes in efficiency of analyte absorption depending on kind of impinger and rate of air stream flow Changes in composition of gaseous sample (combustion gases from incinerator of hazardous waste) Differences in characteristics of indication tubes and hand pumps used during passing gas stream PAH losses from sample trapped on filter depending on volume of air sample passed through filter Permeation of analyte through materials used in construction of sampler (copper, stainless steel, Interferences in quantitative determination when using derivatization technique involving DNPH Changes in cadmium concentrations in aerosol samples depending on particle diameter Effect of impaction, bounce and reaerosolization on collection efficiency of impingers Adsorption of organic compounds from gas phase on particles of suspended phase Effect of rate of gas flow through quartz filter on amount of organic matter trapped Table 11. Sources of error involved in the determination of components in samples of different matrix composition. Sorption of volatile organic compounds on particles of suspended matter Loss of part of aerosols collected by sampler mounted on board plane Description of source of error Escape of some analytes from sample taken on filter or in impactor Sources of error involved in sampling of gaseous components Contamination of sample with combustion gases from turbine Adsorption of gaseous organic components on quartz filter Changes over time in operation efficiency of impingers glass, chromium-plated stainless steel, teflon) tube (DNPH on C₁₈ sorbent) collected in steel canister through tube Sampling of aerosols in upper part of troposphere Sampling of unsaturated aldehydes and ketones Sampling of chlorobenzenes and PCBs from Stage of analytical procedure Sampling of NO₂ from air to impinger Sampling of carbonyl compounds Sampling of HNO₃ from air Sampling of aeroslos Sampling of gases combustion gases Sampling Gaseous Material assayed samples

Talbe 11 (continued)

SOURCE AND A VIOLENCE	Taking and storage of gas samples	Effect of polishing of surfaces of vessels made of PTFE on changes in sample composition	59
Gaseous	Sampling of combustion gases	Effect of devices used on conditions of isokinetic sampling	48
samples	Taking and preparing samples for analysis	Effect of interference (ozone) on determination results of volatile organic compounds	09
	Sampling of dust	Effect of construction material of impactor on results of elemental analysis of dust samples	61
de un romanos abourés nos	Enrichment of moderately volatile organic compounds	Disturbances in recovery of analytes from XAD-2 bed (after extraction stage) due to presence of HCI in combustion gas stream	62
Andrew Market Common	Enrichment of trace amounts of light hydrocarbons from air	Effect of way of preparing sorption tube (filled with several layers of various sorbents) on final determinations	63
	Sampling of analytes to sorption tubes	Analyte losses from sorbent bed due to non-hermetic sealing of tube in storage	64
e'' e sul canadin ministr	Sampling of analytes using high volume air sampler (HVS)	Effect of way of sampler preparation and storage (before use) on results of determination of PCB in air	65
	Sampling of analytes on sorbent hed	Way of purifying sorbent bed before extraction stage	[69-99]
	Camping of araytes of solden bed	Changes in analyte enrichment efficiency depending on sorbent characteristics	70
	Release of components from sorbent bed (thermal desorption)	Dependence of recovery rate of volatile organohalogen compounds on conditions of thermal desorption	71
	Storage of PAH samples on quartz filters	Effect of filter storage conditions on analyte losses	72
	Sampling of analytes from stream of combustion gases on sorbent bed	Effect of HCI presence in gas stream on adsorption of chlorophenols on XAD-2 sorbent bed	62
	Sampling of volatile analytes from gas phase from waste tank	Stability of analyte samples on sorbent bed (after extraction stage)	73
Committee of the Commit	Sampling of analytes from gas stream on sorbent bed	Effect of water vapour on analyte adsorption on carbon bed of molecular sieve	74
	Storage of ozone samples	Instability of absorption solutions (alkaline solution of potassium iodide) after absorption of ozone from air stream	75
		Changes in concentrations of components (H ₂ S, SO ₂ , COS, CH ₃ SH, C ₂ H ₅ SH, CS ₂) of gas samples stored in containers made of Tedlar	76
	Storage of gas samples	Adsorption of organic components on surface of containers made of Tedlar	77
		Changes in composition of gas samples stored in containers made of PVC, Tedlar, polyester, aluminium foil, polyamide, polyethylene	78
	Storage of standard gas mixture	Changes in composition of standard mixture of reactive hydrocarbons (C_2 - C_5) in air stored under pressure	79

	Storage of gas samples and calibration mixtures	Interaction of components of gas mixtures with surface of containers made of aluminium and electropolished stainless steel	80
	Storage of gas samples in plastic containers	Changes in concentrations of volatile organic compounds in gas sample during its storage in containers made of various plastics	81
	Storage of standard gas mixtures	Changes in composition of standard mixtures during their storage	82
	Storage of gas samples	Effect of technique of preparation of teflon surfaces of containers (polishing) on reduction of sample contamination	83
***************************************		Changes in composition of gaseous samples in containers of polished stainless steel	84
	Storage of filters with MDI sample	Changes in stability of sample (diphenylmethane diisocyanate - MDI) trapped on fibreglass filter	85
	Storage of samples in canisters (under pressure)	Changes in humidity of stream of gases released from canister	98
	Storage of sorption tubes	Sunshine- and temperature-induced degradation of α-pinene during storage of tubes with Tenax-TA sorbent (after adsorption stage)	87
	Removal of interfering substances	Changes in composition of gaseous sample depending on ozone concentration; effect of ozone removal technique	92
Gaseous samples	Removal of reactive components from gaseous samples	Effect of scrubber filling on ozone concentration in sample (changes in oxygenation levels of analytes from terpene group	88
	Removal of interferences from gaseous samples	Changes in dimethyl sulfoxide concentration in gaseous sample depending on degree of removal of ${\sf NO}_2$	89
		Change in concentrations of volatile organic analytes in gas stream while drying it in permeation dryer	06
	Sample drying	Effect of dryer (bed of Mg(ClO ₄) ₂) on assays of non-methane hydrocarbons	9
	Extraction of analytes from sorbent bed	Citatige III volume or gas sample due to CO2 ausor priori in bed or sinca ger (or ying substance). Effect of sorbent properties and its level of purity on course of extraction using supercritical fluid	32
	Final determinations	Changes in retention parameters of chromatographic column	94
	Final determinations (GC-ECD)	Changes in response of electron-capture detector	92
	Production of standard gaseous mixtures	Effect of ozone on changes in concentrations of isoprene and methyl vinyl ketone in 'wet' standard gaseous mixture	96
Liquid	Sampling of unsaturated water zone	Dependence of determinations of metallic concentrations on presence of suspension	6
samples	Sampling of rainwater	PCB adsorption from ambient air (collector surface)	98
	Sampling of groundwater	Adsorption of aromatic hydrocarbons on surface of materials used in construction of wells for groundwater sampling (PVC, PTFE, polyethylene, stainless steel, polyvinylidene fluoride epoxyimpregnated fibreglass	66

Talbe 11 (continued)

		Adsorption of pesticides on PVC surfaces	100
	Taking and storage of water samples	Adsorption of organic compounds dissolved in water on PVC and PTFE surfaces	101
		Effect of construction materials of vessels and reagents (acids) used on determination of metal content	102
Liquid samples	Sampling of seawater	Effect of cleanness of sampling system on possibility of determining ultra-trace concentrations (<ppt) lead<="" of="" th=""><th>103</th></ppt)>	103
•	Sampling of water	Effect of amount of matter suspended in water on representativeness of water sample taken to assay metal content in dissolved phase	104
	Taking and storage of seawater samples	Changes in concentration levels of volatile organohalogen compounds	105
	Taking and transport of liquid samples	Leaching of components from polymeric construction materials (FEP, PU, PVC, polyamide)	[106- 107]
	Sampling of water from underground	Effect of sampling technique (piston pump, submersible centrifugal pump) on concentrations of volatile organic compounds (POC)	108
	Storage of analyte (pesticide) samples on bed of sorbent	Effect of storage temperature and level of drying of sorbent bed on stability of herbicide sample	109
	Storage of natural water samples	PAH adsorption on suspension particles, PAH adsorption on walls of vessels	110
	Storage of drinking water (in bottles)	Changes in concentrations of trace components	111
	Storage of standard solutions	Stability of solution components (arsenic compounds)	112
	Storage of groundwater samples	Adsorption of volatile organic compounds on surface of teflon membrane (in sampling chamber located in container stopper	113
	Storage of seawater samples	Effect of sample storage at -20sC on concentration levels of phosphates and nitrates	114
	Storage of water samples	Formation of unstable organophosphates and inorganic polyphosphates in water samples as result of phosphate hydrolysis	115
		Changes in concentration levels of bisphenol A during storage of water samples	116
	Storage of water samples	Effect of way of water sample storage on determination of copper content	117
		Effect of deep freezing of water sample (-70°C) on determination of DOC	118
		Changes in concentrations of nitrites and nitrates in stored water sample. Use of pasteurization to stabilize sample composition	119
	Conservation of water samples	Effect of conserving agent on stability of water samples for assays of non-ionic surfactants	120
		Effect of conserving agent on results of metal content assays	121
	Conservation of seawater samples	Changes in nutrient concentrations in seawater samples during thermal conservation	122

	Extraction of organic analytes from water samples	Effect of conserving agent (HgCl ₂) on efficiency of extraction using solvent and using stream of gas (CLSA)	123
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REFERENCES

- 1. J. Namieśnik, Trends in environmental analytics and monitoring. *Crit. Rev. Anal. Chem.*, **30**, 221–269 (2000).
- 2. J. Namieśnik, Modern trends in monitoring and analysis of environmental pollutants. *Pol. J. Environ. Stud.*, **10**, 127–140 (2001).
- 3. U.A.Th. Brinkman, Multidimensional approaches in environmental analysis. *Anal. Commun.* **34**, 9H-14H (1997).
- 4. O.F.X. Donard, F.M. Martin, Hypenated techniques applied to environmental speciation studies. *Trends Anal. Chem.*, **11**, 17–26 (1992).
- 5. Th.H.M. Noij, F.L. Schulting, Trends in analytical techniques for the analysis and identification of organic micropollutants in water. *Water Supply*, **11**, 31–49 (1993).
- R. Łobiński, Advanced techniques for bioinorganic and environmental speciation analysis for heavy metals. *NATO ASI Ser.*, 26, 389–409 (1997).
- 7. M. Hoenig, Preparation steps in environmental trace element analysis facts and traps. *Talanta*, **54**, 1021–1038 (2001).
- J. Namieśnik, T. Górecki, Preparation of environmental samples or the determination of trace constituents. *Pol. J. Environ. Stud.*, 10, 77–84 (2001).
- J. Namieśnik, T. Górecki, Sample preparation for chromatographic analysis of plant material. *J. Planar Chromatogr.*, 13, 404–413 (2000).
- 10. G. Tölg, Problems and trends in extreme trace analysis for the elements. *Anal. Chim. Acta*, **283**, 3–18 (1993).
- 11. S. Noack, Of what purity are pure substances, *GIT Fachz. Lab.*, **40**, 771–775 (1996).
- 12. W. Büttner, J. Dahmen, N. Harder, J. Heckenkamp, Contamination during the sampling of ultraclean chemicals. **37**, 992–996 (1993).
- 13. M. Gardner, Stability of trace organic substances in water: data obtained from interlaboratory tests. *Anal. Commun.*, **35**, 373–375 (1998).
- 14. M. Feuerstein, G. Schlemmer, B. Kraus, The simultaneous GFAAS determination of

- various elements at ultratrace levels in ultrapure acids and photoresist stripper solutions. *At. Spectrosc.*, **19**, 1–5 (1998).
- G. Knapp, Mechanised methods of sample decomposition in trace and ultra-trace analysis. *Analytical Proceedings*, 27, 112–114 (1990).
- J. Wibowo, F. Shadman, D. Blackford, Measuring and removing dissolved and colloidal silica in ultrapure water. *Micro* (May 1997).
- 17. M. K. Balazs, Ultrapure water, friend or enemy?. *Solid State Technol.*, **36**, 75–81 (1993).
- 18. K. Clark, M. Retzik, D. Darbouret, Measuring TOC to maintain high-purity water. *Ultrapure Water*, 21–24 (February 1997).
- H. Schleibinger, H. Rüden, Air filters from HVAC systems as possible source of volatile organic compounds (VOC) – laboratory and field assays. *Atmospheric Environment*, 33, 4571–4577 (1999).
- A. Lamberty, J. R. Moody, E. van Duffel, P. de Bièvre, J. Broothaerts, Ph. Taylor, C. Lathen, The ultra-clean chemical laboratory (UCCL) at the Institute for Reference Materials and Measurements (IRMM). *Fresenius J. Anal. Chem.*, 357, 359–363 (1997).
- 21. C. P. Ferrari, A. L. Moreau, C. F. Boutron, Clean conditions for the determination of ultra-low levels of mercury in ice and snow samples. *Fresenius J. Anal. Chem.*, **366**, 433–437 (2000).
- 22. C. Aguilar, I. Ferrer, F. Borrull, R. M. Marcé, D. Barceló, Monitoring of pesticides in river water based on samples previously stored in polymeric cartridges followed by on-line solid-phase extraction-liquid chromatographydiode array detection and confirmation by atmospheric pressure chemical ionization mass spectrometry. *Anal. Chim. Acta*, 386, 237–248 (1999).
- 23. G. A. Sorial, M. T. Suidan, R. D. Vidic, R. C. Brenner, Effect of GAC Characteristics on adsorption of organic pollutants. *Water Environ. Res.*, **65**, 53–57 (1993).
- H. Gerstenberger, Zur Durchlässigkeit von Wasser durch die Wandungen von Kunststffflaschen (Water penetration through the walls of containers made from plastics). GIT Fachz. Lab. 4, 279– 284 (1994).

- B. Griepink., Improving the quality of environmental trace analysis. *Fresenius Z. Anal. Chem.*, 317, 210–212 (1984).
- P. Bachhausen, N. Buchholz, H. Hartkamp, Wandeffekte in der Gasspurenanalyse. Fresenius Z. Anal. Chem. 322, 336–342 (1985).
- S. K. Papiernik, A raview of in situ measurement of organic compound transformation in groundwater. *Pest. Manag. Sci.*, 57, 325–332 (2001).
- 28. N. L. Law, M. L. Diamond, The role of organic films and the effect on hydrophobic organic compounds in urban areas: an hypothesis. *Chemosphere*, **36**, 2607–2620 (1998).
- 29. H. M. "Skip" Kingston, D. Huo, Y. Lu, S. Chalk, Accuracy in species analysis: speciated isotope dilution mass spectrometry (SIDMS) exemplified by the evaluation of chromium species. *Spectrochimica Acta B*, **53**, 299–309 (1998).
- G. Benoit, K. S. Hunter, T. F. Rozan, Sources of trace metal contamination artifacts during collection, handling, and analysis of freshwaters. *Anal. Chem.*, 69, 1006–1011 (1997).
- 31. A. E. Childress, M. Elimelech, Effect of solution chemistry in the surface charge of polymeric reverse osmosis and nanofiltration membranes. *J. Membrane Sci.*, **119**, 253–268 (1996).
- 32. M. Hoenig, A.-M. de Kersabiec, Sample preparation steps for analysis by atomic spectroscopy methods: present status. *Spectrochim. Acta B*, **51**, 1297–1307 (1996).
- 33. J. R. W. Woitiez, J. E. Sloof, Sampling and sample preparation. In: Determination of trace elements (Ed.: Z. B. Alfassi), VCH Weinheim (1994) Chapter 3, 59–107.
- 34. J. Yang, T. S. Conver, J. A. Koropchak, D. A. Leighty, Use of a multi-tube Nafion® membrane dryer for desolvation with thermospray sample introduction to inductively coupled plasma-atomic emission spectrometry. *Spectrochim. Acta B*, **51**, 1491–1503 (1996).
- 35. B. King, Development of an international chemical measurement system plenary lecture. *Analyst*, **118**, 587–591 (1993).

- W. Funcke, H. Linnemann, Sampling of polychlorinated dibenzofurans (PCDF) an dibenzo(p)dioxins (PCDD) in emissions from combustion facilities using an adsorption method. *Chemosphere*, 24, 1563–1572 (1992).
- 37. V. L. Foltescu, E. Selin, Corrections for particle losses and sizing errors during aircraft aerosol sampling using a rosemount inlet and the PMS LAS-X. *Atmospheric Environment*, **29**, 449–453 (1995).
- 38. K. G. Heumann, Determination of inorganic and organic traces in the clean room compartment of Antarctica. *Anal. Chim. Acta*, **283**, 230–245 (1993).
- M. Chih Chang, C. Sioutas, S. Kim, H Gong Jr., W. S. Linn, Reduction of nitrate losses from filter and impactor samplers by means of concentration enrichment. *Atmospheric Environment*, 34, 825–98 (2000).
- S. R. McDow, J. J. Huntzicker, Vapor adsorption artifact in the sampling of organics aerosol: face velocity effects. *Atmospheric Environment*, 24A, 2563–2571 (1990).
- 41. C. E. Miller, R. Lewis, The effect of sampling rate on the efficiency of the warren spring sampler in the determination of ambient concentrations of airborne particles. Technical note. *Atmospheric Environment*, **24A**, 2267–2270 (1990).
- 42. Th. W. Kirschstetter, C. E. Corrigan, T. Novakov, Laboratory and field investigation of the adsorption of gaseous organic compounds onto quartz filters. *Atmospheric Environment*, **35**, 1663–1671 (2001).
- 43. U. Knecht, H.-J. Woitowitz, PAH-losses from glass fiber filters under the conditions of different air volume sampling: results of field evaluations in occupational atmospheres. *Fresenius Z. Anal. Chem.*, **331**, 8–13, (1988).
- S. A. Grinshpun, K. Willeke, V. Ulevicius, A. Juozaitis, S. Terzeiva, J. Donnely, G. N. Stelma, K. P. Brenner, Effect of impaction, bounce and reaerosolization on the collection efficiency of impingers. *Aerosol. Sci. Technol.*, 26, 326–342 (1997).
- 45. E. D. Winegar, L. H. Keith, Sampling and analysis of airborne pollutants. 191–208, Lewis Publishers (1993).

- R. Niessner, Sampling techniques for air pollutants. In: *Environmental Analysis* (Ed.: D. Barceló): *Techniques, Applications and Quality Assurance* (Chapter 1), 3–22, Elsevier Sci. Publ., 1993.
- X. Lin, K. Willeke, V. Ulevicius, S. A. Grinshpun, Effect of sampling time on the collection efficiency of all-glass impingers.
 Am. Ind. Hyg. Assoc. J., 58, 480–488 (1997).
- 48. G. Leonardos, F. Sullivan, S. P. Levine, R. T. Stordeur, T. M. Harvey, D. Schuetzle, A comparison of polymer adsorbent and bag sampling techniques for paint bake oven odorous emissions. *J. Air Poll. Control Assoc.*, **30**, 22–29 (1980).
- 49. S. D. Wachasunder, Sh. Satyanarayan, The effect of sample preparation techniques on the measurement of total organic carbon of synthetic waste. *Indian J. Environmental Protection*, **9**, 19–21 (1989).
- 50. W. R. Haag, Interchangeability of gas detection tubes and hand pumps. *AIHAJ*, **62**, 65–69 (2001).
- A. R. Ghoison, J. F. Storm, R. K. M. Jayanty, R. G. Fuerst, T. J. Logan, M. R. Midgett, Evaluation of canisters for measuring emissions of volatile organic air pollutants from hazardous waste incineration. *J. Air Pollut. Control Assoc.*, 39, 1210–1217 (1989).
- 52. K.-U. Goss, S. J. Eisenreich, Sorption of volatile organic compounds to particles from a combustion source at different temperatures and relative humidities. *Atmospheric Environment*, **31**, 2827–2834 (1997).
- 53. G. Desmet, G. Dumont, D. Tielemans, R. de Lathouwer, E. J. Roekens, Measurements of atmospheric pollutants using helicopters: evaluation of the possible contamination of the sample air by turbine exhausts. *Atmospheric Environ.*, **29**, 2547–2552 (1995).
- 54. S. K. Goyal, Measurement of NO₂ in ambient air effect of bubbler type and samole flow rate in sodium arsenite method. *Indian J. Environmental Protection*, **19**, 161–165 (1999).
- 55. Z. Tuba, Z. Csintalan, Z. Nagy, K. Szente, Z. Takács, Sampling of terricolous lichen and moss species for trace element analysis, with special reference to bioindication of air pollution., In: Environmental sampling for trace

- analysis (Ed.: B. Markert), VCH Weinheim (1994) Chapter 22, 415–434.
- F. de Santis, A. Febo, C. Perrino, Negative interference of teflon sampling devices in the determination of nitric acid and particulate nitrate. *Science Total Environ.*, 76, 93–99 (1988).
- 57. M. Pires, L. R. F. Carvalho, An artifact in air carbonyls sampling using C₁₈ DNPH-coated cartridge. *Anal. Chim. Acta*, **367**, 223–231 (1998).
- 58. R. Schulte-Ladbeck, R. Lindahl, J.-O. Levin, U. Karst, Characterization of chemical interferences in the determination of unsaturated aldehydes using aromatic hydrazine reagents and liquid chromatography. *J. Environ. Monit.*, **3**, 306–310 (2001).
- M. Takenaka, M. Hayashi, I. Suzuki, Y. Yamada, K. Takamatsu, M. Kageyama, Evaluation of a mirror-polishing technique for fluorocarbon polymer surfaces for reduction of contamination from containers used in ultratrace analysis. *Anal. Chem.*, 69, 972–976 (1997).
- 60. D. Helmig, Ozone removal techniques in the sampling of atmospheric volatile organic trace gases. *Atmospheric Environment*, **31**, 3635–3651 (1997).
- H. Bayer, A. von Bohlen, R. Klockenkämper, D. Klockow, Choice of a suitable material for construction of a battelle type impactor to minimize systematic errors in sampling of airborne dust. *Mikrochim. Acta*, 119, 167– 176 (1995).
- 62. L. K. Tan, A. J. Liem, Effect of hydrochloric acid on sampling and analysis of semivolatile organic compounds in incineration flue gas. 1. chlorophenols. *Environ. Sci. Technol.*, **30**, 1053–1060 (1996).
- 63. A. Castellnou, N. Gonzales-Flesca, J. O. Grimalt, Refrigerated multibed adsorption in sampling and analysis of atmospheric light hydrocarbons at ppb (v/v) and sub-ppb (v/v) concentrations, *J. Chromatogr. A*, **778**, 269–277 (1997).
- 64. P. J. H. D. Verkoelen, M. W. F. Nielen, Short-term sample loss and sample uptake by Tenax adsorption tubes. *J. High Res. Chromatogr. Chromatogr. Comm.*, **11**, 291–293 (1988).

- I. Basu, J. M. O'Dell, K. Arnold, R. A. Hites, A source of PCB contamination in modified high-volume samplers. *Environ. Sci. Technol.*, 34, 527–529 (2000).
- G. Hunt, N. Pangaro, Potential contamination from the use of synthetic adsorbents in air sampling procedures. *Anal. Chem.*, **54**, 369– 372 (1982).
- 67. N. Schmidbauer, M. Oehme, Comparison of solid adsorbent and stainless steel canister sampling for very low ppt-concentrations of aromatic compounds (≥ C₆) in ambient air from remote areas. *Fresenius Z. Anal. Chem.*, **331**, 14–19 (1988).
- 68. D. Helming, Artifact-free preparation, storage and analysis of solid adsorbent sampling cartridges used in the analysis of volatile organic compounds in air. *J. Chromatogr. A*, **732**, 414–417 (1996).
- S. A. Hubbard, G. M. Russwurm, S. G. Walburn, A method for reducing and evaluating blanks in Tenax air sampling cartridges. *Atmospheric Environ.*, 15, 905–908 (1981).
- I. Devai, R. D. DeLaune, Trapping efficiency of various solid adsorbents for sampling and quantitative gas chromatographic analysis of carbonyl sulfide. *Analytical Letters*, 30, 187–198 (1997).
- 71. U. Herber, H.-U. Meisch, Studies about the desorption of volatile halocarbons from activated carbon by application of static headspace gas chromatography. *Fresenius J. Anal. Chem.*, **353**, 219–221 (1995).
- 72. J. Oda, A. Yasuhara, K. Matsunaga, Y. Saito, Stability of polycyclic aromatic hydrocarbons and their oxygenated derivatives during various storage. *Jpn. J. Toxicol. Environ. Health* **44**, 352–363 (1998).
- C.-Y. Ma, J. T. Skeen, A. B. Dindal, Ch. K. Bayne, R. A. Jenkins, Performance evaluation of a thermal desorption/gas chromatographic/mass spectrometric method for the characterization of waste tank headspace samples. *Environ. Sci. Technol.*, 31, 853–859 (1997).
- M. Gawryś, P. Fastyn, J. Gawlowski, T. Gierczak, J. Niedzielski, Prevention of water vapour adsorption by carbon molecular sieves in sampling humid gases. *J. Chromatogr. A*, 933, 107–116 (2001).

- M. Hekmat, P. Fung, R. Smith, Instability of ozone samples collected in alkaline potassium iodide solution. *Am. Ind. Hyg. Assoc. J.*, 53, 672 (1992).
- Y. K. Lau, Measurement of sulphur gases in ambient air. *Environmental Monitoring and Assessment*, 13, 69–74 (1989).
- 77. L. J. Mc Garvey, Ch. V. Shorten, The effects of adsorption on the reusability of Tedlar® air sampling bags. *AIHAJ*, **61**, 375–380 (2000).
- J. C. Polasek, J. A. Bullin, Evaluation of bag sequential sampling technique for ambient air analysis. *Environ. Sci. Technol.*, 12, 708–712 (1978).
- J. Mowrer, A. Lindskog, Automatic unattended sampling and analysis of background levels of C₂ C₅ hydrocarbons. *Atmospheric Environment*, 25A, 1971–1979 (1991).
- J. J. F. McAndrew, D. Znamensky, D. DeBord, R. Inman, Interaction of gas-phase organic molecules with aluminium and electropolished stainless steel tubing. *Anal. Chem.*, 67, 3075– 3078 (1995).
- 81. J. C. Possner, W. J. Woodfin, Sampling with gas bags I: losses of analyte with time. *Appl. Ind. Hyg.*, **4**, 163–168 (1986).
- R. K. M. Jayanty, J. R. Albritton, Y. H. Straley, D. J. von Lehmden, Stability of PPB organic cylinder gases used for calibration and audits., Proc. 84th Ann. Meet. Exhib., June 16– 21, 1991, Vancouver, BC, Canada, paper 91– 58.2.
- 83. J. H. Shreffler, Comparison of nonmethane organic compound concentration data collected by two methods in Atlanta. *J. Air Waste Manage. Assoc.*, **43**, 1576–1584 (1993).
- 84. K. D. Oliver, J. D. Pleil, W. A. McClenny, Sample integrity of trace level volatile organic compounds in ambient air stored in summa® polished canisters. *Atmospheric Environment*, **20**, 1403–1411 (1986).
- 85. W. J. Karoly, Stability studies of diphenylmethane diisocyanate (MDI) on glass fiber filters. *Am. Ind. Hyg. Assoc. J.* **59**, 645–647 (1998).
- W. A. McClenny, S. M. Schmidt, K. G. Kronmiller, Variation of the relative humidity of air released from canisters after ambi-

- ent sampling. *J. Air & Waste Manage. Assoc.*, **49**, 64–69 (1999).
- 87. W. Schrader, J. Geiger, D. Klockow, E.H. Korte, Degradation of α-pinene on Tenax during sample storage: effects of daylight radiation and temperature. *Environ. Sci. Technol.*, **35**, 2717–2720 (2001).
- 88. J. Fick, L. Pommer, B. Andersson, C. Nilsson, Ozone removal in the sampling of parts per billion levels of terpenoid compounds: an evaluation of different scrubber materials. *Environ. Sci. Technol.*, **35**, 1458–1462 (2001).
- 89. B. A. Rodrigues, L. R. M. Pitombo, A. A. Cardoso, Study on the use of oxidant scrubbers for elimination of interferences due to nitrogen dioxide in analysis of atmospheric dimethylsulfide. *J. Braz. Chem. Sci.*, **11**, 71–77 (2000).
- J. D. Pieil, K. D. Oliver, W. A. McClenny, Enhanced performance of nation dryers on removing water air samples prior to gas chromatographic analysis. *J. Air Poll. Control Assoc.*, 37, 244–248 (1987).
- 91. P. V. Doskey, The effect of treating air samples with magnesium perchlorate for water removal during analysis. *J. High Resol. Chromatogr.* **14**, 724–728 (1991).
- S. W. Klamm, G. W. Scheil, Sample volume measurement errors caused by CO₂ adsorption in desiccants. *Environ. Sci. Technol.*, 23, 1420–1422 (1989).
- 93. J. R. Bowyer, Supercritical fluid extraction as a means of cleaning and desorbing common air sampling sorbents. *Chemosphere*, **31**, 2905–2918 (1995).
- 94. J. B. Laurens, J. M. Swinley, J. P. de Coning, Gas chromatographic analysis of trace impurities in chlorine trifluoride. *J. Chromatogr. A*, **873**, 229–235 (2000).
- 95. J. Lasa, I. Śliwka, Stability extraction of the response of a GC equipped with an ECD working in a constant current mode. *Chem. Anal.* (*Warsaw*), **46**, 421–431 (2001).
- 96. E. Leibrock, J. Slemr, Method for measurement of volatile oxygenated hydrocarbons in ambient air. *Atmospheric Environment*, **31**, 3329–3339 (1997).
- 97. S. E. Bufflap, H. E. Allen, Sediment pore water collection methods for trace metal analysis: a review. *Wat. Res.*, **29**, 165–177 (1995).

- 98. T. J. Murphy, C. W. Sweet, Contamination of teflon surfaces by PCBs in the atmosphere. *Atmospheric Environment*, **28**, 361–364 (1994).
- R. W. Gillham, S. F. O'Hannesin, Sorption of aromatic hydrocarbons by materials used in construction of ground-water sampling wells. *ASTM STP*, **1053**, 108–122 (1989).
- 100. W. C. Koskinen, A. M. Cecchi, R. H. Dowdy, K. A. Norberg, Adsorption of selected pesticides on a rigid PVC lysimeter. *J. Environ. Qual.*, 28, 732–734 (1999).
- L. V. Parker, T. A. Ranney, Effect of concentration on sorption of dissolved organics by PVC, PTFE and stainless steel well casings. GWMR, 139–149 (summer 1994).
- 102. M. Gasparon, Trace metals in water samples: minimizing contamination during sampling and storage. *Environmental Geology*, **36**, 207–214 (1998).
- 103. V. Stukas, C. S. Womg, W. K. Johnson, Subpart per trillion levels of lead and isotopic profiles in a fjord, using an ultra-clean pumping system. *Marine Chemistry*, **68**, 133–143 (1999).
- 104. I. G. Droppo, C. Jaksot, Impact of river transport characteristics on contaminant sampling error and design. *Environ. Sci. Technol.*, **28**, 161–170 (1995).
- 105. J. Pruvost, O. Connan, Y. Marty, P. Le Corre, A sampling device for collection and analysis of volatile halocarbons in coastal and oceanic waters. *Analyst*, **124**, 1389–1394 (1999).
- L. V. Parker, Th. A. Ranney, Sampling tracelevel organic solutes with polymeric tubing. Part I. Static studies. *GWMR*, 115–124 (Fall 1997).
- L. V. Parker, Th. A. Ranney, Sampling tracelevel organic solutes with polymeric tubing. Part II. Dynamic studies. *GWMR*, 148–155 (Winter 1998).
- L. L. Knobel, L. J. Mann, Sampling for purgeable organic compounds using positivedisplacement piston and centrifugal submersible pumps: a comparative study. *Ground Water Monit. Remed.*, 142–148 (spring 1993).
- H. Sabik, R. Jeannot, E. Sauvard, Stability of herbicides and their degradation products on graphitized carbon black extraction cartridges

- used for large volumes of surface water. *Analusis*, **28**, 835–842 (2000).
- 110. H. Zawadzka, J. Zerbe, D. Baratkiewicz, Straty policyklicznychweglowodorów aromatycznych (PWA) podczas przechowywania próbek wód naturalnych., Losses of polycyclic aromatic hydrocarbons (PAH) during storage of natural water samples. *Chem. Anal.*, 25, 469–471(1980).
- 111. R. B. Thurman, S. J. Sterry, How safe is bottled water? *Water*, 7–10 (March/April 1998).
- 112. M. A. Palacios, M. Gómez, C. Cámara, M. A. López, Stability studies of arsenate, monomethylarsonate, dimethylarsinate, arsenobetaine and arsenocholine in deionozed water, urine and clean-up dry residuefrom urine samples and determination by liquid chromatography with microwave-assisted oxidation-hydride generation atomic absorption spectrometric detection. *Anal. Chim. Acta*, 340, 209–220 (1997).
- D. A. Kovacs, D. H. Kampbell, Improved method for the storage of groundwater samples containing volatile organic analytes. *Arch. Environ. Contam. Toxicol.*, 36, 242–247 (1999).
- 114. L. A. Clementson, S. E. Wayte. The effect of frozen storage of open-ocean seawater samples on the concentration of dissolved phosphate and nitrate. *Wat. Res.*, 26, 1171–1176 (1992).
- 115. F. H. Denison, Ph. M. Haygarth, W. A. House, A. W. Bristow, The measurement of dissolved phosphorus compounds: evidence for hydrolysis during storage and implications for analytical definitions in environmental analysis. *Int. J. Environ. Anal. Chem.*, 69, 111–123 (1998).
- 116. D. A. Markham, D. A. McNett, J. H. Birk, G. M. Klecka, M. J. Bartels, Ch. A. Staples, Quantitative determination of bisphenol-A in river water by cool on-column injection-gas chromatography-mass spectrometry. *Int. J. Environ. Anal. Chem.*, 69, 83–98 (1998).
- 117. S. L. Simpson, S. C. Apte, G. E. Batley, Sample storage artifacts, affecting the measurement of dissolved copper in sulfidic waters. *Anal. Chem.*, 70, 4204–4205 (1998).
- 118. K. Hiraki, M. Kimura, H. Hattori, Y. Nakaguchi, H. Tsubota, Determination of

- dissolved organic carbon in Northwest Pacific Ocean. The application of ultra low long time storage. *Kiuki Daigaku Rikogakubu*, **32**, 19–24 (1996).
- 119. L. M. Avalone, D. W. Toohey, W. H. Brune, R. J. Salawitch, A. E. Dessler, J. G. Anderson, Balloon-borne in situ measurements of chlorine oxide and ozone: implications for heterogenous chemistry and mid-latitude ozone loss. *Geophys. Res. Lett.*, 20, 1795– 1798 (1993), Acc. to Chem. Abstr. Vol. 120, 249657s (1994).
- A. Szymański, Z. Zwit, Z. Łukaszewski, Studies of preservation of water samples for the determination of non-ionic surfactants. *Anal. Chim. Acta*, 311, 31–36 (1995).
- J. T. Creed, Th. D. Martin, M. Sivaganesan, Preservation of trace metals in water samples. *Jour. AWWA*, 104–114 (February 1995).
- 122. A. Aminot, R. Kérouel, Assessment of heat treatment for nutrient preservation in seawater samples. *Anal. Chim. Acta*, **351**, 299–309 (1997).
- 123. W. T. Foreman, S. D. Zaugg, L. M. Faires, M. G. Werner, Th. J. Lelker, P. F. Rogerson, Analytical interferences of mercuric chloride preservative in environmental water samples: determination of organic compounds isolated by continuous liquid-liquid extraction or closed—loop stripping. *Environ. Sci. Technol.*, 26, 1307–1312 (1992).
- 124. M. Gardner, S. Comber, Sample filtration as a source of error in the determination of trace metals in marine waters. *Analyst*, **122**, 1029– 1032 (1997).
- 125. M. Gardner, S. Comber, Sample determination of trace metals in marine waters. *Analyst*, **122**, 1029–1032, (1997).
- 126. A. J. Horovitz, Some thoughts on problems associated with various sampling media used for environmental monitoring. *Analyst*, **122**, 1193–1200 (1997).
- 127. A. J. Horovitz, Problems associated with using filtration to define dissolvent trace element concentrations in natural water samples. *Environ. Sci. Technol.* **30**, 954–963 (1996).
- 128. B. Norrman, Filtration of water samples for DOC studies. *Marine Chemistry*, **41**, 239–242 (1993).

- 129. F. Ödman, Th. Ruth, Ch. Pontér, Validation of a field diltration technique for characterization of suspended particulate matter from freshwater. Part I. Major elements. *Applied Geochemistry*, **14**, 301–317 (1999).
- 130. A. J. Horovitz, K. R. Lum, J. R. Garbardino, G. E. M. Hall, C. Lemieux, Ch. R. Demas, The effect of membrane filtration on dissolved trace element concentrations. *Water, Air and Soil Pollution*, **90**, 281–294 (1996).
- 131. D. O. Cooney, Adsorption of organic solutes on membrane filters during aqueous phase filtration. I. Basic rate and equilibrium studies using toluidine blue. *Separation Science and Technology*, **27**, 2001–2019 (1992).
- 132. X. Zhu, M. Elimelech, Colloidal fouling of reverse osmosis membranes: measurements and fouling mechanisms. *Environ. Sci. Technol.*, **31**, 3654–3662 (1997).
- 133. M. Pujo-Pay, P. Raimbault, P. Conan, Sousestimation des teneurs en azote particulaire en milieu océanique par l'utilisation des filtres GF/F. *C. R. Acad. Sci. Paris*, **324**, 401–407 (1997).
- S. C. Yoro, Ch. Panagiotopoulos, R. Semperé, Dissolved organic carbon contamination induced by filters and storage bottles. *Wat. Res.*, 33, 1956–1959 (1999).
- 135. Y. R. Hunter, J. S. Kuwabara, Ionic strength and DOC determinations from various freshwater sources to the San Francisco Bay. *Bull. Environ. Contam. Toxicol.*, **52**, 311–318 (1994).
- 136. A. Sjösten S. Blomqvist, Influence of phosphate concentration and reaction temperature when using the molybdenium blue method for determination of phosphate in water. *Wat. Res.* **31**, 1818–1823 (1997).
- 137. M. Borho, P. Wilderer, A reliable method for preservation and determination of arsenate(III) concentrations in ground water and water works samples. *J. Water SRT – Aqua*, 46, 138–143 (1997).
- 138. E. Turtola, Peroxodisulphate digestion and filtration as sources of inaccuracy in determinations of total phosphorus and dissolved orthophosphate phosphorus in water samples containing suspended soil particles. *Boreal Environment Research*, **1**, 17–26 (1996).

- 139. M. P. Maskarinec, L. H. Johnson, S. K. Holladay, R. L. Moody, Ch. K. Bayne, R. A. Jenkins, Stability of volatile organic compounds in environmental water samples during transport and storage. *Env. Sci. Technol.*, 24, 1665–1670 (1990).
- 140. M. H. Al.-Malack, Migration of lead from unplasticized polyvinyl chloride pipes. *Journal of Hazardous Materials*, **B 82**, 263–274 (2001).
- 141. M. J. Barcelona, J. A. Helfirch, E. E. Garske, Sampling tubing effects on groundwater samples. *Anal. Chem.*, **57**, 460–464 (1985).
- 142. R. Hopman, T. J. J. van den Hoven, Permeation of organic chemicals through plastic water pipes. *J. Water SRT Aqua*, **41**, 158–162 (1992).
- 143. J. Zerbe, D. Baratkiewicz, H. Gramowska, J. Żminkowska, Efektywność144. ekstrakcji policyklicznych węglowodorów aromatycznych z wód o dużej zawartości zawiesin. *Chem. Anal.*, 29, 455–459 (1984).
- 144. M. Lyytikäinen, J. Pellinen, Some issues concerning the gas chromatographic determination of chlorinated phenolics in water. *Toxicological and Environmental Chemistry*, **63**, 185-197 (1997).
- 145. M. Palma, Z. Piñeiro, C. G. Barroso, Stability of phenolic compounds during extraction with superheated solvents. *J. Chromatogr. A*, **921**, 169–174 (2001).
- 146. I. Ferrer, D. Barceló, Determination and stability of pesticides in freeze-dried water samples by automated on-line solid-phase extraction followed by liquid chromatography with diode-array detection. *J. Chromatogr. A*, 737, 93–99 (1996).
- 147. G. A. Sorial, M. T. Suidan, R. D. Vidic, R. C. Brenner, Effect of GAC characteristics on adsorption of organic pollutants. *Water Environ. Res.*, **65**, 53–57 (1993).
- 148. B. Maichin, P. Kettish, G. Knapp, Investigation of microwave assisted drying of samples and evaporation of aqueous solutions in trace element analysis. *Fresenius J. Anal. Chem.*, **366**, 26–29 (2000).
- P. J. Cerminara, G. A. Sorial, S. P. Papadimas,
 M. T. Suidan, M. A. Moteleb, Th. F. Speth,
 Effect of influent oxygen concentration on

- the GAC adsorption of VOCs in the presence of BOM. *Wat. Res.*, **29**, 409–419 (1995).
- E. Diamadopoulos, P. Samaras, G. P. Sakellaropoulos, The effect of activated carbon properties on the adsorption of toxic substances. *Wat. Sci. Tech.*, 25, 153–160 (1992).
- P. H. Chen, W. A. Van Ausdale, W. S. Keeran,
 D. F. Roberts, GC/MS identification of artifacts formed during sample preparation using
 USEPA methods 625 and 8270. *Chemosphere*,
 26, 1743–1749 (1992).
- 152. M. Vogel, W. Pötter, U. Karst, Characterization of a chemical artifact in the liquid chromatographic determination of 3–butyn-2–one using the 2, 4–dinitrophenylhydrazine method. *J. Chromatogr. A*, **886**, 303–307 (2000).
- 153. W. D. Bowers, M. L. Parsons, R. E. Clement, F. W. Karasek., Component loss during evaporation-reconstruction of organic environmental samples for gas chromatographic analysis. *J. Chromatogr.*, **207**, 203–211 (1981.
- 154. D. E. Slauenwhite, P. J. Wangersky, Extracton of marine organic matter on XAD-2: Effect of sample acidification and development of an in situ preacidification technique. *Mar. Chem.*, 54, 107–117 (1996).
- 155. J. Kukkonen, J. F. McCarthy, A. Oikari, Effects of XAD-8 fractions of dissolved organic carbon on the sorption and bioavailability of organic micropollutants. *Arch. Environ. Contam. Toxicol.*, 19, 551–557 (1990).
- 156. W. E. Johnson, N. J. Fendinger, J. R. Plimmer, Solid-phase extraction of pesticides from water: possible interferences from dissolved organic material. *Anal. Chem.*, 63, 1510–1513 (1991).
- 157. M. Johansson, H. Emteborg, B. Glad, F. Reinholdsson, D. C. Baxter, Preliminary appraisal of a novel sampling and storage technique for the speciation analysis of lead and mercury in seawater. *Fresenius J. Anal. Chem.* 351, 461–466 (1995).
- 158. B. Kolb, G. Zwick, M. Auer, A water trap for static cryo-headspace gas chromatography. *J. High Resol. Chromatogr.*, **19**, 37–42 (1996).
- 159. F. Jüttner, B. Meon, O. Köster, Quasi in situ separation of particulate matter from lakewater by hollow-fibre filters to overcome errors

- caused by short turnover times of dissolved compounds. *Wat. Res.*, **31**, 1637–1642 (1997).
- 160. H. R. Rogers, Influence of suspended solids and back diffusion on organic contaminant uptake by semi-permeable membranes (SPDMs). *Chemosphere*, 35, 1651–1657 (1997).
- 161. C. Molina, P. Grasso, E. Benfenati, D. Barceló, Determination and stability of phenmediphan, ethofumesate and fenamiphos in ground water samples using automated solid phase extraction cartridges followed by liquid chromatography high flow pneumatically assisted electrospray mass spectrometry. *Int. J. Environ. Anal. Chem.*, 65, 69–82 (1996).
- 162. K. Dettmer, Th. Knobloch, W. Engewald, Stability of reactive low boiling hydrocarbons on carbon based adsorbents typically used for adsorptive enrichment and thermal desorption. *Fresenius J. Anal. Chem.*, 366, 70–78 (2000).
- 163. H. Emteborg, J. Snell, J. Qian, W. Frech, Sources of systematic errors in mercury speciation using Grignard reagents and capillary gas chromatography coupled to atomic spectrometry. *Chemosphere*, 39, 1137–1152 (1999).
- T. J. Nestrick, L. L. Lamparski, Purification of cylinder gases in solvent evaporation for trace analysis. *Anal. Chem.* 53, 122–124 (1981).
- 165. N. A. Stevens, M. F. Borgerding, Optimization of ¹⁵N detection with an Atomic Emission Detector. *Anal. Chem.*, 71, 1083–1086 (1999).
- 166. T. Ozeki, Y. Tsubosaka, S. Nalayama, N. Ogawa, T. Kimoto, Study of errors in determination of hydrogen ion concentrations in rainwater samples using glass electrode method. *Anal. Sci.*, 14, 749–756 (1998).
- 167. Ch. Weston, D. Speis, R. P. Albert, A comparison of the performance characteristics of sorbent column packing materials used for purgeable organic analysis. Technical information, Environmental Testing and Certification Corp., Edison, New Jersey, 1991.
- 168. K. E. Collins, C. H. Collins, C. A. Bertran, Stainless steel surfaces in LC systems, part I – corrosion and erosion. LCGC, 18, 600–608 (2000).

- 169. K. E. Collins, C. H. Collins, C. A. Bertran,
 Stainless steel surfaces in LC systems, part II
 passivation and practical recommendations.
 LCGC, 18, 688–692 (2000).
- 170. S. Blomqvist, S. Westin, Interference from chromate, germanate, tungastate and vadnadate when determining phosphate in aqueous solution by the phosphoantimonylomolybdenum blue method. *Anal. Chim. Acta*, **3580**, 245–254 (1998).
- 171. S. Blomqvist, K. Hjellström, A. Sjösten, Interference from arsenate, fluoride and silicate when determining phosphate in water by the phosphoantimonylomolybdenum blue method. *Intern. J. Environ. Anal. Chem*, **54**, 31–43 (1993).
- 172. L. Wennrich, J. Efer, W. Engewald, Gas chromatographic trace analysis of underivatized nitrophenols. *Chromatographia*, **41**, 361–366 (1995).
- 173. J. Poehlman, B. W. Pack, G. M. Hieftje, A heated transfer line for coupling GC with plasma source spectrometry. *Am. Lab.*, 50C-53C (Oct. 1998).
- 174. M. J. Lichon, Environmental monitoring problems with stream sampling. *Water*, **1–2**, 30–32 (1996).
- 175. I. Héninger, M. Potin-Gautier, I. de Gregori, H. Pinochet, Storage of aqueous solutions of selenium for speciation at trace level. *Fresenius J. Anal. Chem.*, 357, 600–610 (1997).
- 176. R. Muñoz Olivas, P. Quevauviller, O. F. X. Donard, Long term stability of organic selenium species in aqueous solutions. *Fresenius J. Anal. Chem.*, **360**, 512–519 (1998).
- 177. J. Snell, J. Qian, M. Johanssos, K. Smit, W. Frech, Stability and reactions of mercury species in organic solution. *Analyst*, **123**, 905–909 (1998).
- 178. D. Amoroux, Ch. Pécheiran, O. F. X Donard, Formation of volatile selenium species in synthetic seawater under light and dark experimental conditions. *Appl. Organometal. Chem.*, **14**, 236–244 (2000).
- 179. W. Chatkittikunwong, C. S. Creaser, Stability of bromo- and bromochloro-dibenzo-p-dioxins under laboratory and environmental

- conditions. *Chemosphere*, **29**, 547–557 (1994).
- 180. G. E. M. Hall, J. C. Pelchat, G. Gauthier, Stability of inorganic arsenic(III) and arsenic(V) in water samples. *J. Anal. At. Spectrom.*, **14**, 205–213 (1999).
- 181. F. P. L. Lamé, P. R. Defize, Sampling of contaminated soil: sampling error in relation to sample size and segregation. *Environ. Sci. Technol.*, **27**, 2035–2044 (1993).
- 182. R. Djingova, I. Kuleff, On the sampling of vascular plants for monitoring of heavy metal pollution. In: *Environmental Sampling for Trace Analysis* (Ed.: B. Markert), VCH Weinheim (1994) Chapter 21, 395–414.
- 183. R. S. Daniels, A. L. Fraser, Ph. Quevauviller, The variability if dry-mass correction for certified reference materials. *J. Environ. Anal. Chem.*, **63**, 167–177 (1996).
- 184. U. Faure, P. J. Wagstaffe, Stability of reference materials. *Fresenius J. Anal. Chem.*, **345**, 124–126 (1993).
- 185. S. Vandendriessche, B. Griepink, J. C. Th. Hollander, J. W. J. Gielen, F. G. G. M. Langelaan, K. J. Saunders, R. H. Brown, Certification of a reference material for aromatic hydrocarbons in Tenax samplers. *Analyst*, 116, 437–441 (1991).
- 186. D. van Dijk, V. J. G. Houba, Homogeneity and stability of materials distributed within the wageningen evaluating programmes for analytical laboratories. *Commun. Soil Sci. Plant. Anal.*, **31**, 1745–1756 (2000).
- 187. P. V. Doskey, R. W. Talbot, Sediment chronologies of atmospheric deposition in a precipitation-dominated seepage lake. *Limnol. Oceanogr.*, **45**, 895–904 (2000).
- 188. D. Huo, H. M. "Kip" Kingston, Correction of species transformations in the analysis of Cr(VI) in solid environmental samples using speciated isotops dilution mass spectrometry. *Anal. Chem.*, 72, 5047–5054 (2000).
- 189. S. B. Hawthorne, C. B. Grabanski, Vaporization of polycyclic aromatic hydrocarbons (PAHs) from sediments at ambient conditions. *Environ. Sci. Technol.*, **34**, 4348–4353 (2000).
- 190. I. T. Cousins, H. Kreibich, L. E. Hudson, W. A. Lead, K. C. Jones, PAHs in soils: contem-

- porary UK data and evidence for potential contamination problems caused by exposure of samples to laboratory air. *Sci. Total. Environ.*, **203**, 141–156 (1997).
- 191. H. Hintelmann, Comparison of different extraction techniques used for methylmercury analysis with respect to accidental formation of methylmercury during sample preparation. *Chemosphere*, **39**, 1093–1105 (1999).
- 192. L.-O. Kjeller, Ch. Rappe, Soxhlet extraction parameters' influence on the recovery of polychlorinated dibenzo-p-dioxins and dibenzofurans (PCDD/F) from a sediment and artifact formation of PCDD. *Toxicol. Environ. Chem.*, 67, 481–490 (1998).
- 193. K. Takada, T. Ashino, K. Wagatsuma, Development of a nitrogen atmosphere pretreatment chamber for the determination of trace amounts of carbon in high-purity iron by the infrared absorption method after combustion in an electric resistance furnace. *Analytical Sciences*, 13, 867–871 (1997).
- S. J. Lue, C. Huang, Applications of ion chromatography in the semiconductor industry.
 II. Determination of basic airborne contaminants in a clean room. *J. Chromatogr. A*, 850, 283–287 (1999).
- 195. P. Sun, M. Adams, L. Shive, S. Pirooz, Molecular and ionic contamination monitoring for cleanroom air and water surfaces. *SPIE*, **3215**, 118–127 (1997).
- 196. P. A. Tanner, L. S. Leong, The effects of different drying methods for marine sediment upon moisture content and metal determination. *Marine Pollution Bulletin*, 31, 325–329 (1995).
- 197. F. Bordas, A. C. M. Bourg, A critical evaluation of sample pretreatment for storage of contaminated sediments to be investigated for the potential mobility of their heavy metal load. *Water*, *Air and Soil Pollution* **103**, 137–149, (1998).
- 198. M. Siaka, C. M. Owens, G. F. Birch, Evaluation of some digestion methods for the determination of heavy metals in sediment

- samples by flame-AAS. *Analytical Letters*, **31**, 703–718 (1998).
- 199. G. Stringari, I. Pancheri, F. Möller, O. Failla, Influence of two grinding methods on the uncertainty of determinations of heavy metals in atomic absorption spectrometry/electrothermal atomization of plant samples. *Accred. Qual. Assur.*, 3, 122–126 (1998).
- 200. M. Takenaka, S. Kozuka, M. Hayashi, H. Endo, Determination of ultratrace amounts of metallic and chloride ion impurities in organic materials for microelectronics devices after a microwave digestion method. *Analyst*, 122, 129–132 (1997)
- 201. Y. Cai, M. Abalos, J. M. Bayona, Effects of complexing agents and acid modifiers on the supercritical fluid extraction of native phenyl- and butyl-tins from sediment. *Appl. Organometal. Chem.*, 12, 577–584 (1998).
- I. L. Marr, P. Kluge, L. Main, V. Margerin, C. Lescop, Digests or extracts? Some interesting but conflicting results for the widely differing polluted sediment samples. *Mikrochim. Acta*, 119, 219–232 (1995).
- 203. L. Jorhem, Dry ashing, sources of error and performance evaluation in AAS. *Mikrochim Acta*, **119**, 211–218 (1995).
- I. Novozamsky, H. J. van der Lee, V. J. G. Houba, Sample digestion procedures for trace element determination. *Mikrochim. Acta* 119, 183–189 (1995).
- B. A. Schumacher, S. E. Ward, Quantitation reference compounds and VOC recoveries from soils by purge-and-trap GC/MS. *Environ. Sci. Technol.*, 31, 2287–2291 (1997).
- 206. L.-O. Kjeller, Addition of internal standards to particulate sample matrices for routine trace analyses of semivolatile organic compounds: a source of systematical and random errors. *Fresenius J. Anal. Chem.*, 361, 791–796 (1998).
- 207. C. Sarzanini, E. Mentasti, Removal of instrumental interferences in evaluating solid waste interaction with soil. *Ann. Chim.*, **85** 407–418 (1995).